Safety & Environmental Assurance Centre

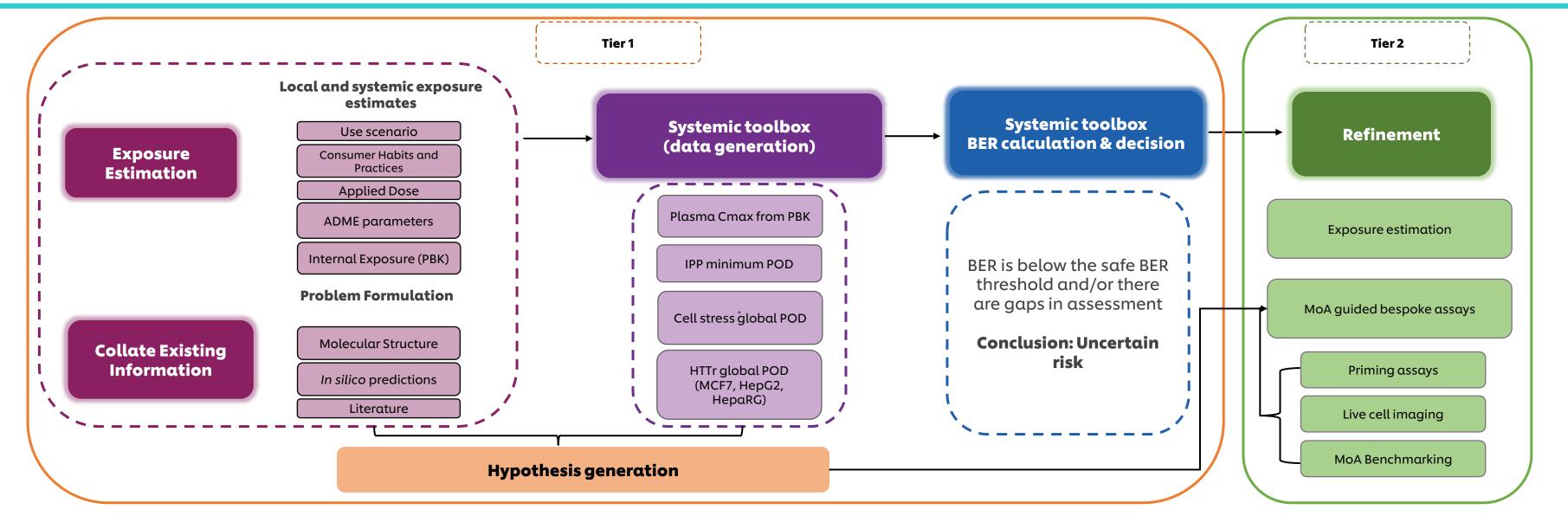


A Next Generation Risk Assessment (NGRA) case study for the bioactive food component sulforaphane

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Introduction

We previously developed a workflow for a 'tier 1' systemic toxicity assessment based on integrating in vitro points of departures (PoDs) from a cell stress panel (CSP), in vitro pharmacological profiling (IPP) and high-throughput transcriptomics (HTTr) with Physiologically-based Kinetic (PBK) Modelling predictions of human exposure to calculate a bioactivity exposure ratio (BER)¹ [Fig 1]. This approach shows promise in a capacity that protects consumers, however, it may be conservative, as PoDs are based on bioactivity in in vitro assays which may not necessarily translate into adverse effects in humans, and many substances (especially food ingredients), display bioactivity at consumer relevant exposures.



This was exemplified for sulforaphane (SFN), a component of cruciferous vegetables [Fig 2]. Numerous studies have linked *Cruciferae* intake with beneficial effects e.g., decreased risk of cancer, with SFN widely hypothesised as a plausible agent for this protection². Given dietary exposure to SFN may have benefits, it is unsurprising that bioactivity occurs at equivalent *in vitro* exposures, illustrating a challenge for the assessment of bioactive substances under the current NGRA paradigm.

Under scenarios where the tier 1 safety assessment cannot enable a safety decision, a tier 2 assessment may be required to elucidate whether the bioactivity would ultimately cause adaptative or adverse effects in humans. The composition of such an assessment is bespoke, however is informed by the hypothesised mode of action (MoA) indicated through tier 1 testing and pre-existing literature knowledge [Fig 1].

In this study, we have conducted a tier 2 hypothetical assessment for SFN to inform a safety decision, focusing on the potential for systemic toxicity using 2 different SFN exposure scenarios that are known to be low risk to humans.

Figure 1. Tier 1 safety assessment framework implemented in previous evaluation and possible areas for refinement in the context of data available for SFN as part of a tier 2 assessment using NAMs¹.

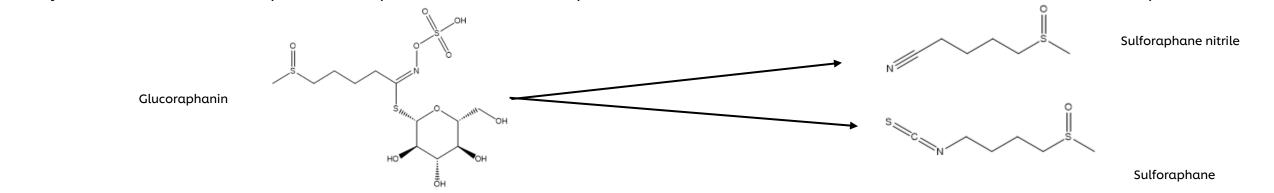
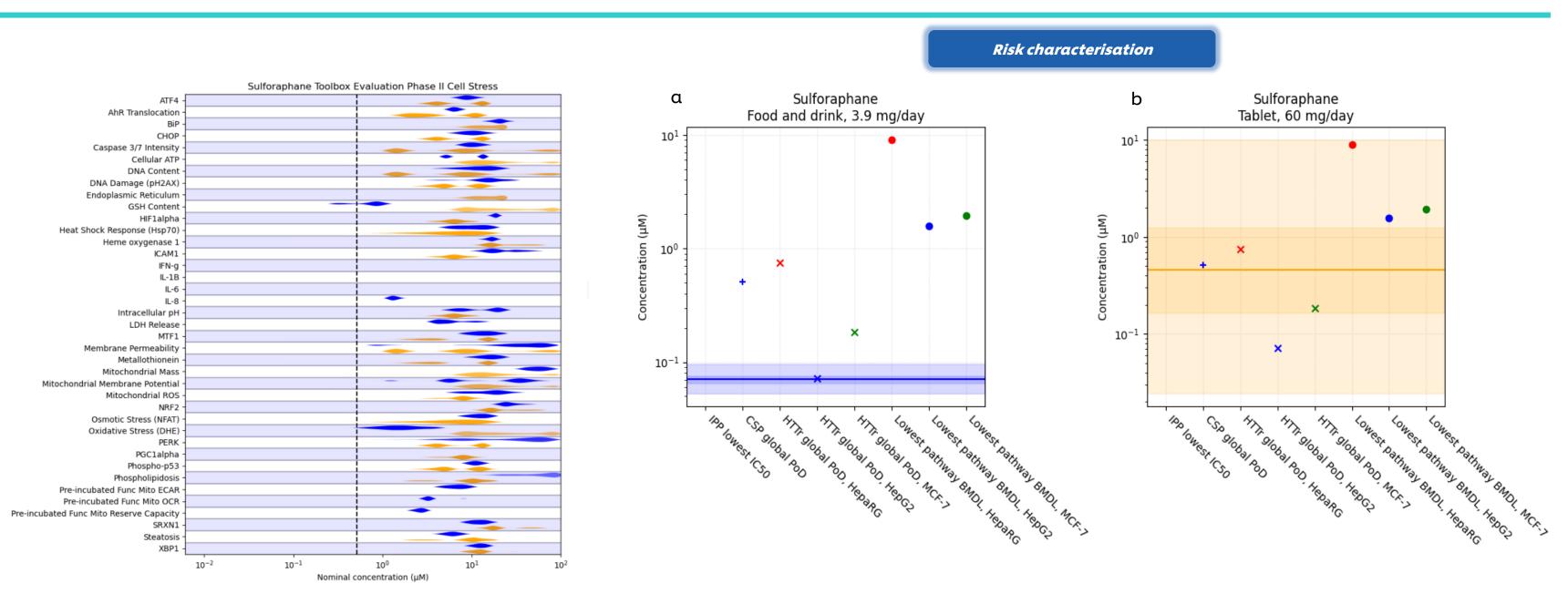


Figure 2. Simplified sulforaphane formation process in cruciferous vegetables. Sulforaphane is not found at considerable concentrations in unprocessed Cruciferae. Instead is stored as a precursor, glucoraphanin. Upon damage to the plant, glucoraphanin is converted to an unstable intermediate (not shown) via the enzymatic action of myrosinases, either present in the plant or in the gastrointestinal tract. This unstable intermediate is converted to sulforaphane or sulforaphane nitrile, with the proportion of conversion to either depending on e.g., temperature, pH etc.



Tier 1 data

	External exposure and risk classification	Internal exposure
Exposure Estimation	Two 'low risk' exposure scenarios were selected for sulforaphane ¹ :	vitro PoDs, external
	Scenario 1: 3.9 mg/day (oral) – Considered representative of 'normal' consumption of Broccoli.	exposures are converted to internal exposures (as Cmax) using PBK Modelling [Fig 4].
	Scenario 2: 60 mg/day (oral) – A clinical trial comprising intake of SFN at 20 mg 3 x daily.	
	IPP	CSP
	SFN showed no hits at a screening concentration of 10 μ M.	Across the CSP, several
	SFN showed no hits at a screening concentration of 10 $\mu\text{M}.$ HTTr	Across the CSP, several biomarkers were perturbed at concentrations prior to



Figure 3. Summary of CSP bioactivity for Sulforaphane. Blue densities indicate PoDs for assay-specific biomarkers and orange densities indicate pooled PoDs for assay-specific cell health biomarkers. Vertical line at 0.51 µM represents the best estimate/'Global PoD' across all biomarkers (corresponding to change in GSH content)⁴

Figure 4. PoDs from Tier 1 in vitro assays and internal exposure (Cmax) estimates (blue/orange line) for both SFN scenarios. A.) for food and drink exposure scenario (3.9 mg/day) b.) for 6-month clinical study involving intake of 60 mg/day SFN. Blue and orange lines in a/b represent exposure estimates with shaded areas representing uncertainty in Cmax estimate, with darker shaded area representative of 50th percentile and lighter area representative of 95th percentile^{1,5}.

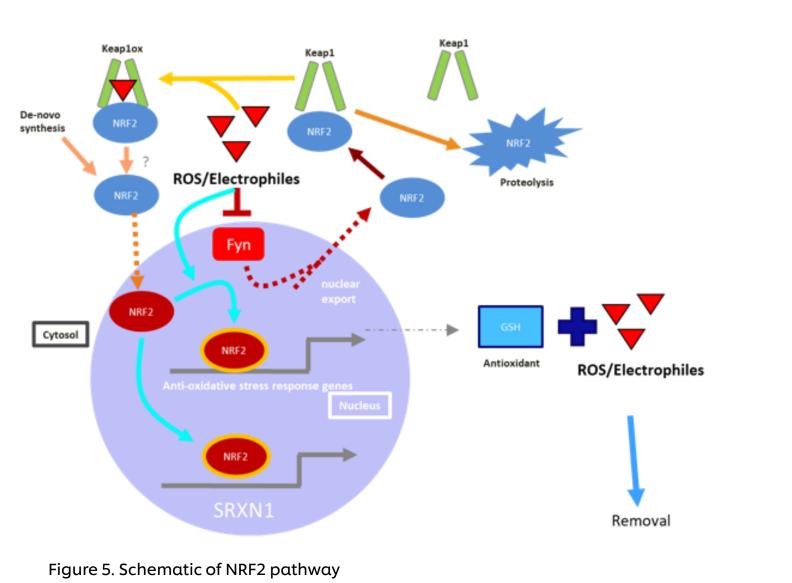
Literature knowledge:

- Soft electrophiles such as SFN form covalent adducts with nucleophiles of similar softness, for example cysteine residues on proteins and GSH.
- One protein susceptible to SFN is KEAP1, the negative regulator of Nrf2 [Fig 5]. Nrf2 is responsible for the transcriptional regulation of >200 genes containing the antioxidant response element (ARE) which have a range of functions such as redox balance/inflammation [Fig 5].
- Although Nrf2 induction by SFN has a potentially beneficial, cytoprotective impact against sources of oxidative stress, at higher exposures, soft electrophiles are well known for causing GSH depletion, oxidative stress and consequently cytotoxicity⁶.

Tier 1 data:

Tier 2 data

• The above information is supported by tier 1 data e.g., in silico profiling (not shown), where positive alerts were returned for protein binding and from the bioactivity assays e.g., the CSP [Fig 4], where the lowest PoDs primarily relate to GSH depletion and reactive oxygen species (ROS) accumulation (eventually causing cytotoxicity).



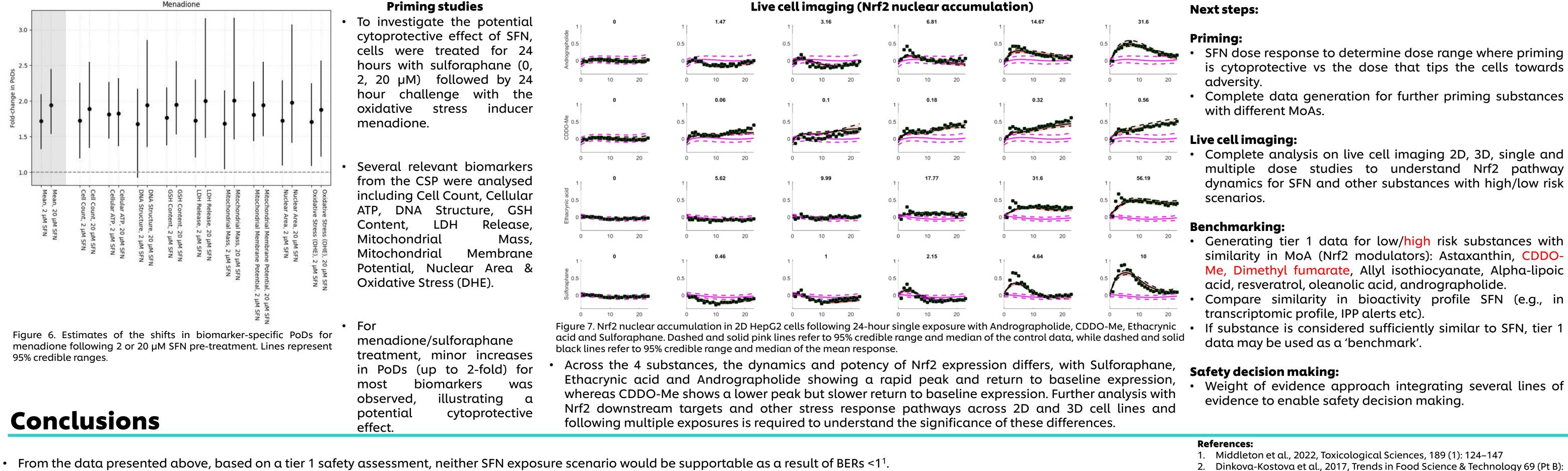
Tier 2 testing strategy:

data/literature informed hypothesis, tier Based on characterisation of SFN's effect on the oxidative stress pathway is necessary to identify a 'tipping point' between adaptation and adversity. Further assays investigating this include:

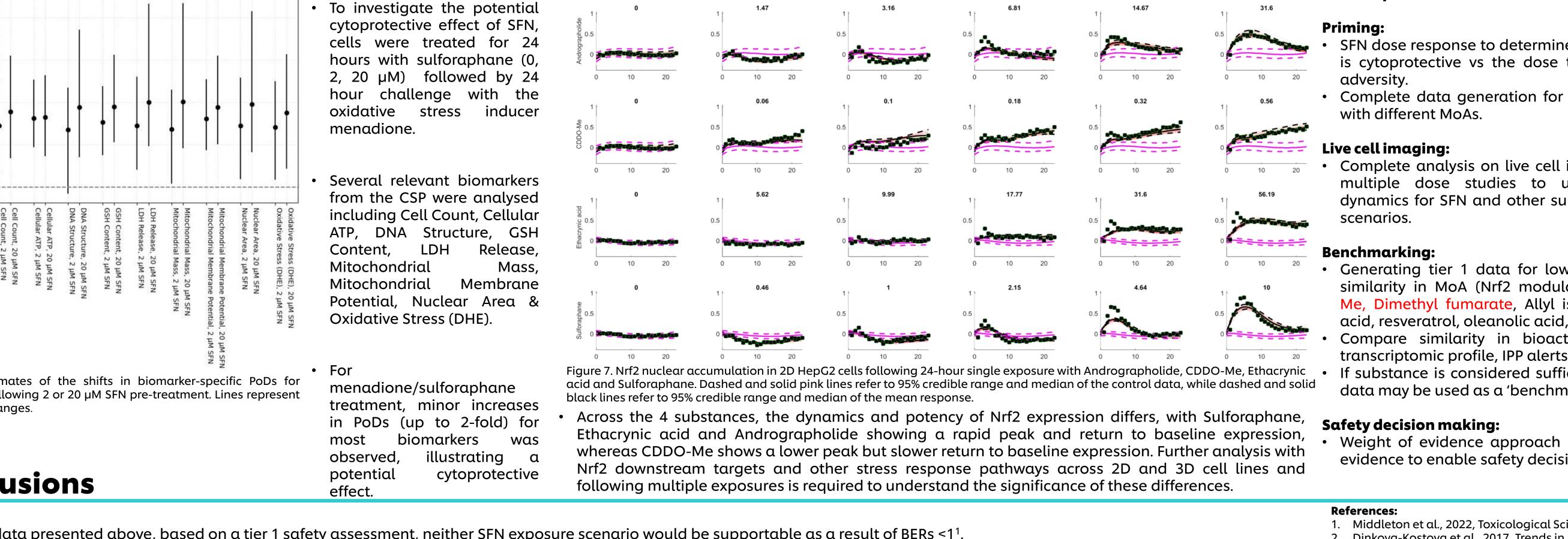
- Priming assays: Pre-treatment of cells with SFN prior to challenge with an oxidative stress inducer (menadione). Analysis to compare SFN + menadione PoDs with menadione only PoD [Fig 6].
- Live cell imaging: Live monitoring of Nrf2 pathway dynamics after single/repeat dosing in 2D and 3D cells with SFN and 'similar' substances (e.g., CDDO-Me, Andrographolide, ethacrynic acid) [Fig 7].
- Benchmarking: Comparing tier 1 bioactivity profile for SFN to other substances with similar MoAs.
- Pathway analysis: Understanding toxicological relevance of differentially expressed genes at consumer relevant exposures.

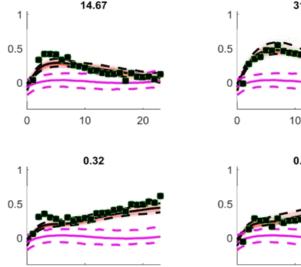
from the BIFROST method at 0.072 µM (HepG2)^{1,3}.

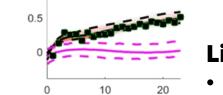
Tier 2 hypothesis generation



Priming studies







257-269

SFN dose response to determine dose range where priming is cytoprotective vs the dose that tips the cells towards

Complete data generation for further priming substances

Complete analysis on live cell imaging 2D, 3D, single and multiple dose studies to understand Nrf2 pathway dynamics for SFN and other substances with high/low risk

- Such a decision from tier 1 does not mean that an exposure scenario constitutes a safety risk per se, instead, SFN-induced bioactivity is predicted following such exposures. Under such scenarios, a tier 2, bespoke assessment is required to understand whether the bioactivity predicted would ultimately cause adaptative or adverse effects in humans.
- The areas of focus for a tier 2 assessment will be case-dependent and could focus on either refining exposure estimates (e.g., through generating further in vitro ADME data or generating human PK clinical data) or refining the hazard characterisation element, where follow-up assays will depend on the bioactivity/hypothesised MoA concluded following tier 1 testing/review.
- For SFN, from the tier 1 data and literature knowledge, further characterisation of the oxidative stress response was chosen as the bioactivity area to focus on.
- Several tier 2 approaches are under way, including priming studies, pathway analysis, live cell imaging, and benchmarking studies. Following the completion and analysis of these studies, a weight of evidence decision may be possible to enable safety decision making.
- 6. LoPachin et al., 2019, Toxicology, 418: 62-69 SAFETY SCIENCE IN THE 21ST CENTURY For more information visit https://seac.unilever.com/

Reynolds et al., 2020, Computational Toxicology, 16, 100138

Hatherell et al., 2020, Toxicological Sciences, 176 (1): 11-33

5. Li et al. 2022, Toxicology and Applied Pharmacology, 442.