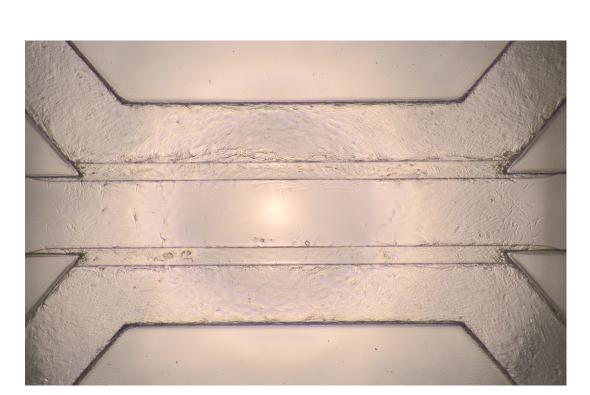






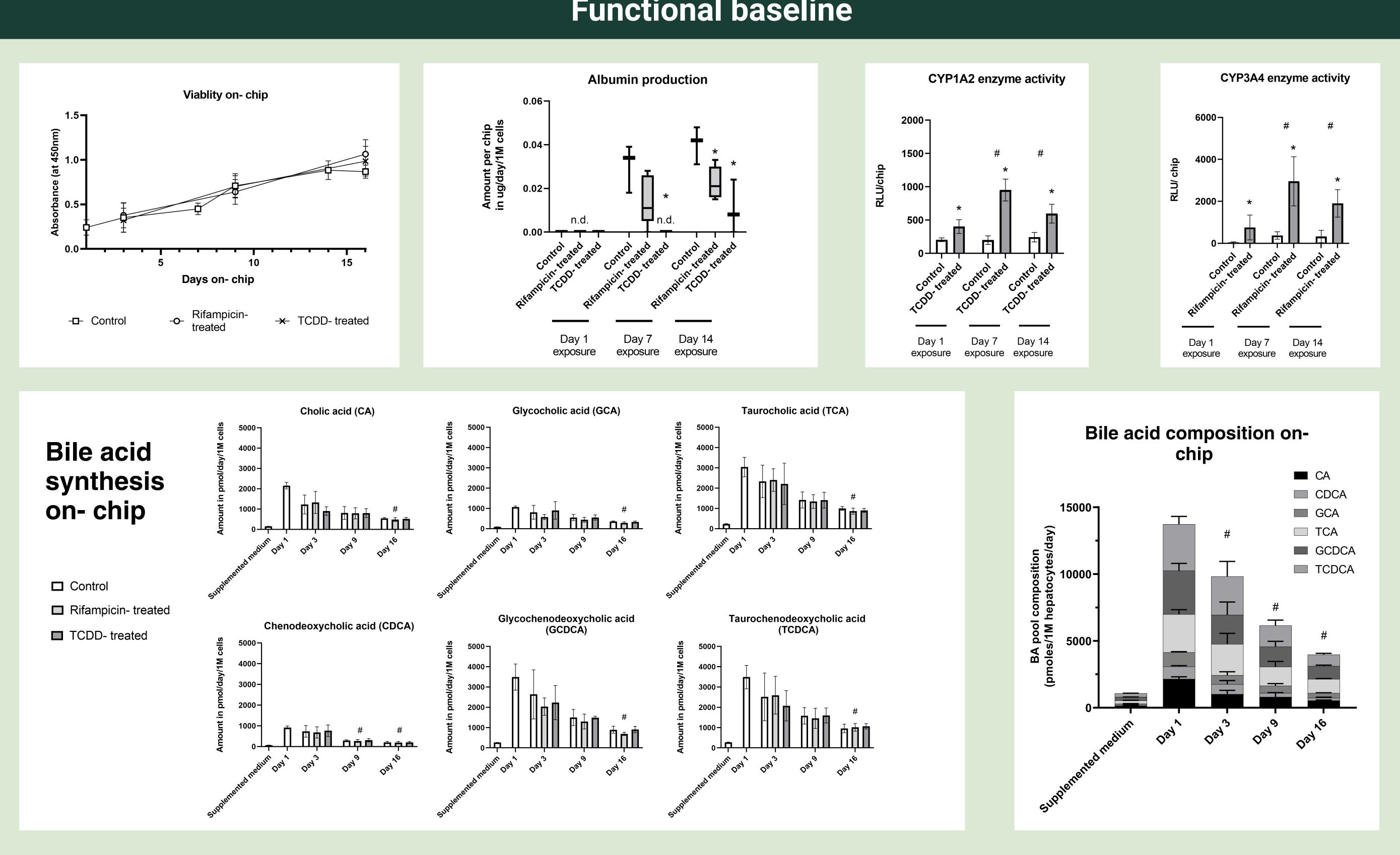
Liver- on- chip



Bright- field image of the liver 3- lane on the tissue Organoplate (Mimetas B.V.)

Liver models are required to evaluate for chemical **biomodulation and biotransformatio**n, as well as for **mechanism- based hepatoxicity** studies¹. Within a Next-Generation Risk Assessment toolbox, Organ- on- chip systems offer the potential to generate data which can be used in a higher tier approach for **biokinetic refinements, targeted** biological mechanism testing and point of departure estimation².

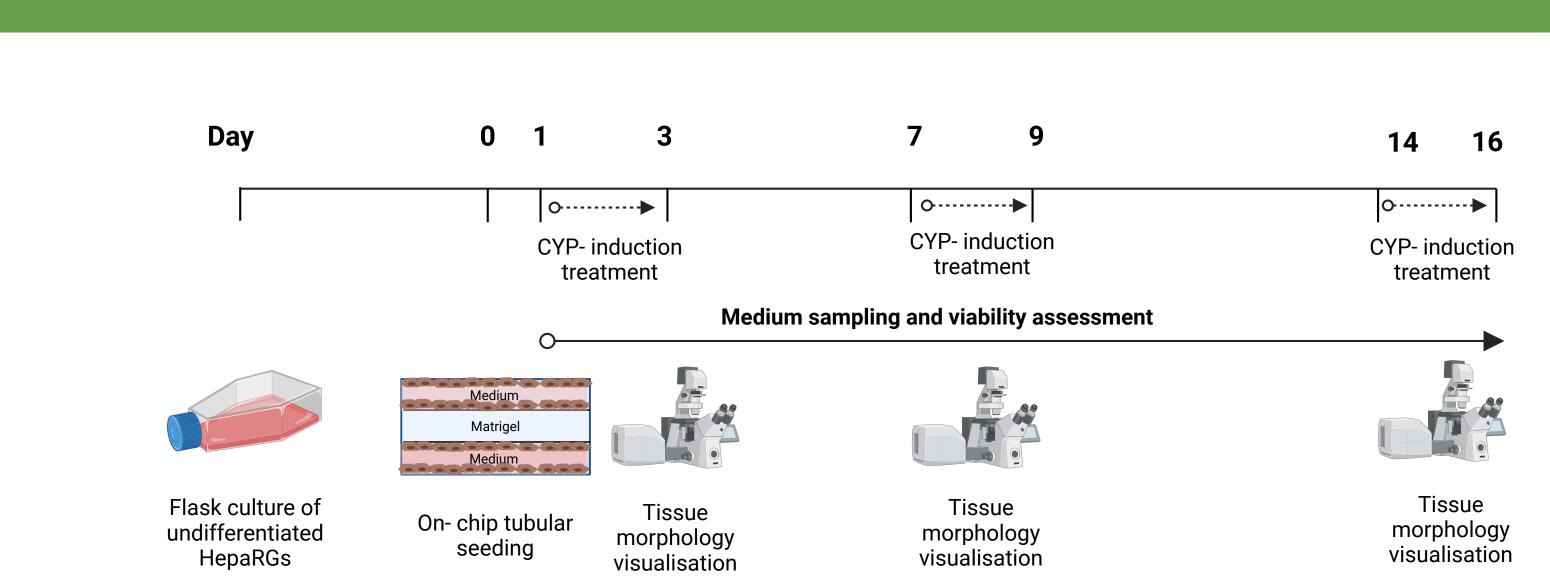
This study aimed to assess **liver- relevant functional baseline** markers for this medium- throughput system at different time points after a **DMSO- free differentiation on- chip** to evaluate the suitability as a cholestasis model.



independent experiments.

A liver- on- chip to evaluate bile acid secretion for the use in a Next- Generation Risk Assessment

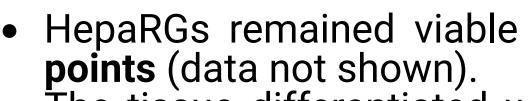
Katharina S. Nitsche¹, Iris Mueller², Wouter Bakker¹, Paul Carmichael^{1,2} and Hans Bouwmeester¹ ¹Division of Toxicology, Wageningen University, P.O. box 8000, 6700 EA Wageningen, the Netherlands ²Unilever Safety and Environmental Assurance Centre, Colworth Science Park, Sharnbrook, Bedfordshire, MK44 1LQ, UK



Functional baseline

Functional measurements cell viability (WST- 8 assay), albumin production (ELISA), CYP enzyme activity (Promega P450 Glo) and bile acid synthesis (LCMS) of the HepaRG liver on- chip to evaluate the model robustness at different time points (n=3). * n.d. = not detected (< LOD); Values represent the mean ±SD of triplicate measurements of at least three

Differentiation and treatment on- chip



- day on-chip (see CYP induction)
- set-up.
- treatment after 7 days on- chip.
- decreased over time.
- not demonstrate a treatment effect
- The bile acid pool decreased overall but remained at the same composition ratios

Collectively, the data demonstrates that HepaRGs on- chip produce an *in vivo* like bile acid profile but also that more culture set- up refinement is needed to increase the functional baseline as a fit- for- purpose cholestasis model in a **Next- Generation Risk Assessment toolbox.**

Acknowledgements

I am thankful for the personal grant for my research project from Unilever SEAC and the supervision from Paul Carmichael, Iris Mueller and Sophie Malcomber. I am grateful to everyone from WUR-TOX for the scientific discussions, as well as the University Fund Wageningen which contributed to my travel to the SOT.

> UNIVERSITY FUND WAGENINGEN



Overview of differentiation and treatment on- chip. HepaRG cells (Biopredic) were precultured in- flask and seeded against Matrigel in the medium channels of the 3lane Organoplate (Mimetas B.V.). The liver-tissue was treated for 48h for CYP enzyme induction with Rifampicin or TCDD on either Day 1, Day 7 or Day 14. For all cultures, the tissue morphology was visualised, the medium sampled and the viability assessed on Day 1, as well as on the exposure start and end days to determine secretion profiles and cytotoxicity.

Results summary

• HepaRGs remained viable on- chip but entered the Matrigel at early time

• The tissue differentiated without dimethyl sulfoxide (DMSO) within the first

• Albumin production increased over the duration of culture. However, the tissue produces only a fraction of albumin compared to other reported perfused liver- models, suggesting a too high shear stress for this seeding

• Under treatment with TCDD and Rifampicin (hepatotoxins known to alter protein biosynthesis), the **albumin production declined as expected**. • Metabolic competency for CYP1A2 and 3A4 was the highest for induction

• A substantial amount of glycine- and tauro conjugated bile acids was de novo synthesised, resulting in a human-comparable liver and bile profile, given that 30% are tauro- conjugates. The perfused model produced also more bile acids compared to reported static models³, even though levels

• De novo bile acid synthesis decreased with the duration of culture and **did**



